




Review Article

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The Human Microbiome: An Overview of Microbial Mechanisms, Diagnostics, and Therapeutic Modulation in Cancer

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Abstract

The human microbiome consists of trillions of microbes (microorganisms) residing in various locations in the body, but primarily in the gut. It plays a vital role in health maintenance and influences several diseases, including cancer. Emerging evidence suggests that microbial dysbiosis, characterized by an imbalance in microbial numbers and diversity, may contribute to cancer development through several mechanisms, including the release of genotoxins, altered metabolism, immune system modulation, and chronic inflammation. This narrative review compiles existing insights into the interactions between the microbiome and cancer, with a particular emphasis on microbial mechanisms during carcinogenesis and the contributions of the microbiome to the diagnosis and treatment of cancer. We examine the potential use of prebiotics, probiotics, fecal microbiota transplants, and microbial therapies that may influence cancer and its response to treatment. Gaining a deeper understanding of the intricate relationship between microbial communities and cancer could lead to groundbreaking methods for cancer prevention and therapy.

Keywords: Cancer, Carcinogenesis, Human microbiome, Microbiota.

الميكروبيوم البشري نظرة عامة على الآليات المايكروبية والتشخيص والتعديل العلاجي في السرطان

الخلاصة

يتكون الميكروبيوم البشري من تريليونات الكائنات الحية الدقيقة التي تعيش في مواقع مختلفة من الجسم، وخاصة في الأمعاء. ويغلب دورًا حيويًا في الحفاظ على الصحة، ويؤثر على العديد من الأمراض، بما في ذلك السرطان. وتشير الأدلة الناشئة إلى أن خلل التوازن الميكروبي، الذي يتميز باختلال التوازن في أعداد الميكروبات وتنوعها، قد يساهم في تطور السرطان من خلال آليات متعددة، بما في ذلك إطلاق السموم الجينية، واضطرابات التمثيل الغذائي، وتعديل الجهاز المناعي، والالتهاب المزمن. تجمع هذه المراجعة الرؤية المتاحة حول التفاعلات بين الميكروبيوم والسرطان، مع التركيز على الأبحاث المهمة التي تربط بين المجموعات المايكروبية وأنواع مختلفة من السرطان. وتتناول الاستخدام المحتمل للميكروبيوم في تشخيص وعلاج السرطان، بما في ذلك زرع ميكروبات البراز والعلاجات المايكروبية التي قد تؤثر على السرطان واستجابته للعلاج. إن فهم العلاقة المعقدة بين المجتمعات المايكروبية والسرطان قد يؤدي إلى أساليب رائدة للوقاية من السرطان وعلاجه.

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INTRODUCTION

Of the estimated 10^{12} different microbial species on earth, the human body is colonized by a relatively small number of approximately 3×10^3 species, including bacteria, viruses, archaea, and unicellular/multicellular eukaryotes, collectively known as “microbiota” [1,2]. In line with scientific rigor and clarity, the term “microbiota” refers to the collection of microorganisms, while the microbiome encompasses not only the microorganisms but also their genomes and metabolites. Bacteria are as numerous as human body cells, and both are estimated to be around 38 trillion (3.8×10^{13}) [3]. The gene count of the human-colonizing microbes exceeds our genome by about 10-fold, enabling diverse metabolic functions and effects on the host, making a human being a ‘hybrid organism’ [4]. These microbes colonize every part of our body surface in contact with the environment, such as the gut, respiratory tract, urogenital tract, skin, etc., from birth and persist until death (Figure 1). Most of the

bacteria inhabiting our body (97% of them) are in the colon, and the remainder are extracolonic. Intratumoral bacteria may also be present at an estimated average number of 0.68%, with a wide range from individual tumors exhibiting no bacteria while others contain nearly 70% bacteria by cell count [5].

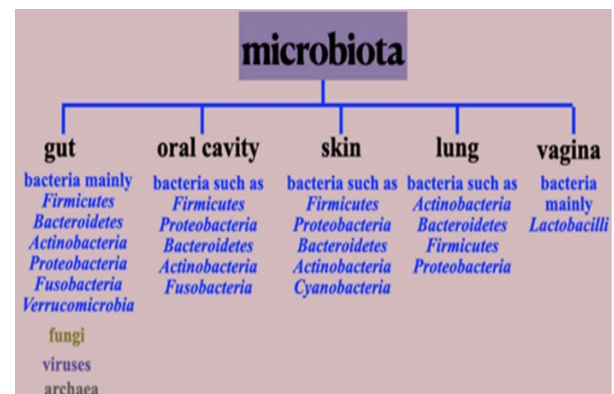


Figure 1: A general outline of the human microbiota and the habitat of its main constituents.

Both human-infecting viruses and bacteriophages may even be present in greater numbers than bacteria [6]. The focus in this narrative review will be “the bacteriome.” The most microbially populated part of the human body is the gut, with the colon making up around 70% of that population [4]. The estimated surface area of the gut is about 200 m², which is equivalent to the area of a tennis court enriched with the required nutrients for the microbes. The bacterial content of the gut increases on travelling from the stomach to the colon, with the stomach being inhospitable to the survival of many species, containing 10 bacterial cells/g, the duodenum (10³), the jejunum (10⁴), the ileum (10⁷), and the colon (10¹²) [7]. Accordingly, the gut represents a major part of the body inhabited by microbes in numbers, varieties, and influences. Although the microbes that inhabit our body establish a beneficial relationship, it is also clear that an imbalance between the types and numbers of the microorganisms can develop. This imbalance, termed dysbiosis, can lead to diseases including cancer (Figure 2) [8]. This review aims to highlight the contribution of microbiomes to carcinogenesis and the influence it might exert on cancer treatments.

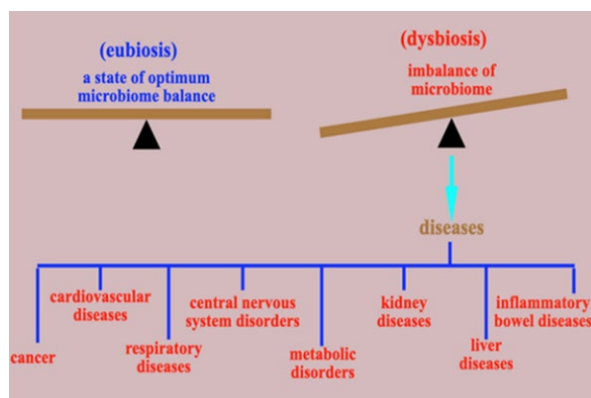


Figure 2: Imbalance of the microbiota in the body can shift the balance from the normal state of eubiosis to a diseased state of dysbiosis.

Location and Composition of the Human Microbiota

Of the large number of species of microorganisms that inhabit our bodies, only 11 of them are recognized to be carcinogenic by the International Agency for Research on Cancer (IARC) [9,10]. The list includes 7 viruses, 3 flatworms, and only one bacterium, with the conspicuous absence of members of the mycobiome (species of fungi). The human virome (the entire population of viruses in a person) is composed of 98% bacteriophages (viruses that are associated with and infect bacteria) and 2% eukaryotic viruses [11]. The seven carcinogenic viruses listed in group 1 of the IARC classification include Epstein-Barr virus (EBV, causing Burkitt lymphoma), hepatitis B virus (HBV, causing hepatocellular carcinoma), hepatitis C virus (HCV, causing hepatocellular carcinoma), Kaposi sarcoma herpesvirus (KSHV, causing Kaposi sarcoma), human immunodeficiency virus-1 (HIV, causing Kaposi sarcoma and non-Hodgkin lymphoma), human papillomavirus (HPV, causing cervical cancer), and

human T-cell lymphotropic virus type 1 (HTLV, causing T-cell malignancies). One additional virus associated with Merkel cell carcinoma (Merkel cell virus –MCV) is listed in group 2A of the IARC list as probably carcinogenic. The three carcinogenic flatworms on the IARC’s list are *Opisthorchis viverrini* (associated with cancer of the gallbladder and its ducts), *Clonorchis sinensis* (associated with bile duct cancer), and *Schistosoma haematobium* (associated with cancer of the urinary bladder). The only bacterial species on the IARC’s list is *Helicobacter pylori* (associated with stomach cancer). However, more recent studies suggest that several other bacteria could modulate or contribute to carcinogenesis, including *Fusobacterium nucleatum*, *Escherichia coli*, *Bacteroides fragilis*, and *Salmonella enterica* [1]. The usual habitat of microbiota, the range of microorganisms inhabiting the body, is the mucosal lining of the gut, skin, oral cavity, respiratory tract, and urogenital tract (see Figure 1). However, in addition to the mucosal microbial niche, intratumoural microbes form a distinct ecosystem that may have an impact on carcinogenesis, and their functions are only beginning to be explored [1,12]. The intratumoural microbiome promotes the progression of tumors by inducing genomic instability and mutations that can lead to alterations in the epigenome, immune/inflammatory response, and the activation of invasion and metastasis. The gut microbiota is considered the most important for health, with significant variations among individuals comparable to fingerprints [4,13]. Gut bacteria have several functions, including the stimulation of the immune response, vitamin production, fermentation of food, and the protection against pathogens [13]. The composition of the microbiota in a newborn infant resembles that of the vagina if the mode of delivery was through cesarean section and matures after three years [8]. Following that, the composition remains stable throughout adulthood unless disease onset, antibiotic use, or change of diet alters that norm [8].

Microbial Mechanisms Driving Carcinogenesis

The mechanisms by which the microbiome can influence cancer initiation and progression in the host fall into three broad categories: a) changing cell proliferation and death, b) changing the immune system function, and c) altering the metabolism. The initiation and promotion of carcinogenesis are largely dependent on the type and species of the cohabiting microbe, in addition to its interaction with the host microenvironment. Oncogenic viruses can start and spread cancer by encoding proteins that control gene expression, cell growth, cell death, and blood vessel formation, as well as immune and inflammatory responses [14]. The mycobiome, the fungal component of the microbiome, can influence carcinogenesis in several ways, including genome instability, inflammation, immunosuppression, and generally through interactions with bacterial residents [15]. The parasitic members of the microbiome may also induce carcinogenesis through the secretion of substances that

can lead to malignant transformation and/or trigger chronic inflammatory response and immune function alterations [16]. Despite the importance of the virome and the mycobiome, we will restrict our discussion to the detailed mechanisms of the bacterial content of the microbiome. Bacteria can promote cancer development and drug resistance through well-documented mechanisms that are generally classified into seven broad categories: 1) increasing genomic damage, 2) impairing DNA repairs, 3) manipulating signalling pathways, 4) altering host metabolism, 5) influencing host immunity, 6) increasing inflammation, and 7) increasing resistance to therapies [17]. There may well be substantial interconnections between these mechanisms in cancer development. Figure 3 illustrates the mechanisms by which the bacteria residing in the gut can contribute to the development of cancer and response to therapies.

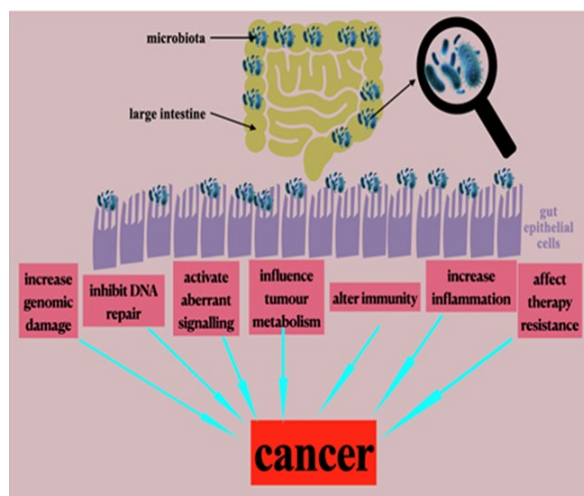


Figure 3: Mechanisms illustrating the influence of the gut bacteria in driving carcinogenesis.

Increasing genomic damage

Direct interactions of bacterial structural components and their secondary metabolites can cause host DNA mutation [18]. *H. pylori*, which colonizes the gastric mucosa of half of the world's population, can cause chronic gastric inflammation, which can progress to gastric cancer in about 1-3% of the colonized individuals [8]. The mechanism is largely attributed to the release of cytotoxin-associated gene (CagA) and the secretion of virulence factors such as VacA, urease, and NapA2 to cause host DNA damage and promote chronic inflammation and oxidative stress [19,20]. Chronic inflammation caused by the infection can also promote host genomic DNA damage, as illustrated by the fact that mice chronically infected with *H. pylori* show a 4-fold rise in mutation frequency compared to uninfected mice [21]. The polyketide synthase, *pks*, pathogenic island, which is carried by certain pathogenic strains of *E. coli*, encodes the genotoxin colibactin. Colibactin creates a crosslink between DNA strands by alkylating the DNA molecule on adenine residues. This causes double-strand breaks, which stops the cell cycle, causes senescence, and encourages the growth of cancer [17,22]. This bacterium, *E. coli*, along with several other bacterial species such as

Campylobacter jejuni, possesses another family of enotoxins called cytolethal distending toxins (CDTs) that cause extensive DNA damage [17,20].

Impairing DNA repair

Cells frequently use DNA mismatch repair (MMR), a significant and well-studied process, to repair genes and preserve the genome. *E. coli* and *H. pylori* deplete these specialized proteins, such as MSH2 and MLH1, in cultured colon cells, thus facilitating mutagenesis [23]. The bacterium *F. nucleatum* provides a different mechanism to deplete the MMR proteins in that it increases the expression of miR-205-5p, which suppresses MSH2, MLH1, and MSH6 regulation, leading to DNA damage and cell proliferation in head and neck squamous cancer [24]. Deficient MMR activity results in a hypermutated phenotype characterized by microsatellite instability (MSI), which is prevalent in around 15% of colorectal cancers.

Manipulating signaling pathways

The cytotoxin-associated gene (CagA), already mentioned above, produced by *H. pylori* can activate β -catenin, leading to the upregulation of genes associated with gastric cancer. E-cadherin/ β -catenin signaling is also turned up by substances made by other bacteria, like *S. typhimurium* and *F. nucleatum*, which are known as AvrR and FadA, respectively [25]. The AvrR protein is also inserted into host cells during *Salmonella* infections to activate JAK/STAT and β -catenin/Wnt signaling, ultimately leading to cancer [26]. *Salmonella typhimurium* infection particularly activates MAPK/Akt signaling, which is essential in the maintenance of gallbladder cancer in mouse models of this disease [27]. The binding of FadA, secreted by *F. nucleatum*, to E-cadherin inhibits this protein's tumor suppressor activity [8]. *Porphyromonas gingivalis* promotes cell proliferation in colorectal cancer through the activation of the MAPK/ERK signalling pathway [28]. Dysregulation of tumor suppressor proteins such as p53, PTEN, and pRb by certain microbes can induce STAT3 signaling, leading to enhanced epithelial-mesenchymal transition and senescence [17].

Altering host metabolism

Bacteria can regulate the glycolysis of the immune cells through toll-like receptor (TLR) signaling. When TLR was activated in macrophages, for instance, the glycolysis was significantly raised, thus increasing the expression of the glucose transporter, GLUT1, and the enzymes hexokinase and enolase [29]. The increased TLR signaling also induces a metabolic shift from oxidative phosphorylation to glycolysis. Bacteria such as *Bacteroides uniformis* have a strong capacity for glycolysis and the production of butyrate, which accumulates in cancer cells due to the Warburg Effect, acting as inhibitors of histone deacetylation [30]. The inhibition of histone deacetylation induces apoptosis and suppresses cell proliferation in colorectal cancer [31]. About 95% of the bile acids that are released into the intestine are reabsorbed through the portal vein and

return to the liver to be recycled. This hepatic-intestinal cycle takes place several times a day (roughly about 6 times a day in humans) [17]. Bile acids can serve as signalling molecules through binding to receptors such as the farnesoid X receptor (FXR), predominantly found in the nucleus of liver and ileum cells. These receptors play a role in the production, transportation, and usage of triglycerides and the metabolism of other lipid molecules [32].

Influencing host immunity

Bacteria and the molecules associated with them activate and recruit innate cells such as NK cells, neutrophils, macrophages, myeloid-derived suppressor cells, and innate lymphoid cells. Additionally, cells from the adaptive immune system, including T and B cells, influence changes in the immune system [33]. The killing ability of NK cells is directly inhibited by the presence of *F. nucleatum* in the TME [34]. This inhibition is partly mediated by the binding of the bacterial Fap2 protein to the human TIGIT receptor. Moreover, increased presence of *F. nucleatum* in the colorectal tumor microenvironment is associated with a reduced number of CD3⁺T cells, which are known to have favorable clinical outcomes [35]. Fap2 is overexpressed in inflamed colon and particularly enriched in colonic adenomas (benign) and adenocarcinoma (malignant) tissues, where it can exist bound to the carbohydrate molecules Gal-GalNac (a disaccharide often used as a tumour marker) [36]. Bacteria can also influence response to immunotherapy, as in the case of several commensals such as *Bifidobacterium longum*, *Collinsella aerofaciens*, and *Enterococcus faecium*, which were found to yield better outcomes to anti-PD1 therapy when present [34]. *Helicobacter pylori* bacteria activate TLR2 and TLR4, causing profound effects on the host immune response through the secretion of inflammatory cytokines such as IL-1 β and IL-18 [37].

Increasing inflammation

Mouse models of carcinogenesis generally revealed that the gut epithelium with a higher density of polyps has a higher permeability compared to healthy tissues, thus allowing more transmucosal bacterial translocation [38]. These translocated bacteria then induce the release of pro-inflammatory cytokines such as IL-6, IL-11, IL-17, IL-22, and IL-23, which participate in cancer progression [20,38]. Short-chain fatty acids (SCFAs), particularly butyrate and propionate, can downregulate the expression of pro-inflammatory cytokines [39]. It is thought that *H. pylori* affects the growth of cancer mainly through inflammatory pathways that are controlled by neutrophils and lymphocytes and boosted by cytokines like IL-1 β , TNF α , and IFN γ [9]. FadA secreted by *F. nucleatum* binds to epithelial and endothelial cells, allowing cellular internalization and the induction of pro-inflammatory cytokines like IL-6 and NF- κ B that can result in oral squamous cell carcinoma [40]. Enterotoxigenic *B. fragilis* is associated with colitis and colon cancer through its secreted toxin, BFT [41].

This toxin stimulates inflammation in colorectal cancer by targeting the tight junctions of cells in the intestine, cleaving E-cadherin, and increasing permeability and inflammation [9].

Increasing resistance to therapy

Resistance to cancer therapy can arise from interactions between the microbiota and the intended therapy, influencing drug metabolism and/or host immunity. The microbiota, and their derivatives, can break down medicines and directly convert them into nontoxic metabolites (or with reduced toxicity) [42]. For example, the bacteria *Mycoplasma hyorhinis* have many enzymes, such as thymidine phosphatase, which break down pyrimidine nucleoside analogues, a class of drugs resembling the building blocks of DNA or RNA that are used to treat cancer and various infections [43]. The same bacteria also possess cytidine deaminase and pyrimidine nucleoside phosphorylase, which can reduce the toxicity (in this case, a desired biological effect) of the chemotherapy drug gemcitabine. Bacteria can also mediate host tolerance to chemotherapies. For instance, irinotecan (a topoisomerase 1 inhibitor employed for several cancers) is converted to the metabolite 7-ethyl-10-hydroxycamptothecin (SN-38) by the intestinal bacteria. The active metabolite, SN-38, is then inactivated by glucuronidation to its inactive form, SN-38G [44]. Certain species of *Clostridium* convert the inactive form, SN-38G, back again to the active SN-38, resulting in severe diarrhea [17]. Ionizing radiation, when used as anticancer therapy, is partially mediated by the stimulation of the major arms of the immune system, innate and adaptive immunity. The main reason for tolerance against this form of therapy is attributed to the disruption of the gut bacteria [17]. Gram-positive bacteria reduce the activity of dendritic cells and other antigen-presenting cells, limit CD8⁺T cell activity, and weaken radiation-induced antitumor response. Vancomycin was shown to improve the radiation-mediated anticancer effect by eliminating gram-positive bacteria [45]. Not all bacteria have a negative influence on cancer therapies. It was found that vitamin B6 produced by *E. coli* is needed for fluorouracil efficacy to be realized in the model organism *C. elegans* [46]. Certain species of *Enterococcus* might elevate intratumoral CD8⁺/Treg cell ratio, thus permitting a more immunosupportive environment.

Contributions of the Microbiome to the Diagnosis and Therapy of Cancer

The microbiota inhabiting our bodies could influence the diagnosis and management of cancer in three ways: 1) in diagnosis, 2) in modulating the effects of other cancer treatment modalities, and 3) in driving new treatment options (Figure 4).

The Microbiome in the Diagnosis of Cancer

The main criteria for a good diagnostic method for the detection of cancer are being sensitive, specific, non-invasive, and able to discover malignancy early enough

to institute preventative and/or treatment measures. Three main options for sampling are available: 1) through the saliva, 2) through the fecal material, and 3) through the plasma for DNA.

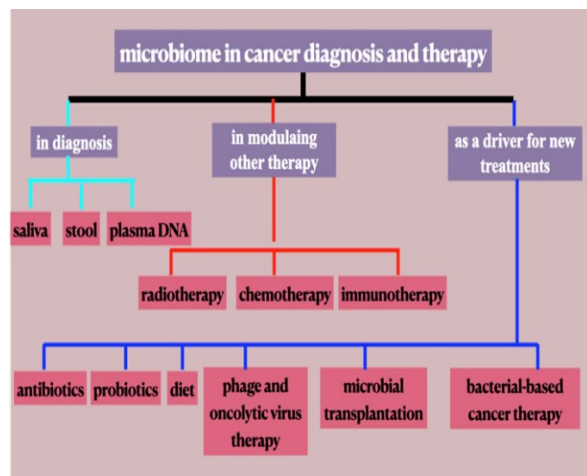


Figure 4: Applications of the human microbiome in cancer diagnosis and therapy.

Through salivary sampling

A combination of three bacteria, *Capnocytophaga gingivalis*, *Prevotella melaninogenica*, and *Streptococcus mitis*, was found to be a good diagnostic indicator in patients with oral squamous cell carcinoma (OSCC) compared to controls, with specificity and sensitivity more than 80% [47]. Saliva samples with *Neisseria elongata* and *Streptococcus mitis* yielded a specificity of 82% and a sensitivity of 96% in patients with pancreatic cancer [48]. In patients with squamous cell lung cancer, a combination of two bacterial genera, *Capnocytophaga* and *Veillonella*, served as a diagnostic indicator with 87% specificity and 85% sensitivity compared with controls [49]. The salivary microbiome sampling is non-invasive and easy to collect, but a lack of collection standards may be one of its main drawbacks.

Through fecal (stool) sampling

Here, most of the supporting research focused on colorectal cancer. A stool sample having a combination of *Lachnospirillum* species with *F. nucleatum*, *Hungatella hathewayi*, and *Bacteroides clarns* yielded a specificity of 81% and a sensitivity of 94% [50]. Accuracy and consistency across populations remain challenges that require further investigation.

Through Plasma DNA (liquid biopsy)

The microbes might leave DNA fragments indicating their presence in different tissues. Circulating pieces of these DNAs in the plasma could act as a useful diagnostic indicator. A next-generation sequencing analysis revealed that these DNA fragments relate to bacteria, though some were also from fungi and viruses [51]. Chen and colleagues developed a model for the early diagnosis of lung cancer and for the prediction of disease recurrence after surgery based on circulating microbial DNA [52]. This diagnostic test yielded a

sensitivity of 87.7% and achieved an area under the curve (a measure of the overall accuracy in distinguishing between diseased and healthy individuals) of 93.2%. Moreover, this test demonstrated the capability to detect early lung cancer with a sensitivity of more than 86%. Although plasma DNA analysis can be a cost-effective diagnostic tool for lung cancer, the method requires further investigation to be applied across many cancers.

The microbiome in modulating cancer therapies

The microbiome was found to be associated with the effects of different forms of cancer therapies, including 1) radiotherapy, 2) chemotherapy, and 3) immunotherapy.

Radiotherapy

The gut microbiome can influence how patients respond to radiotherapy, affecting both its effectiveness and side effects. Eradication of Gram-positive bacteria using antibiotics improved the antitumor effects of radiation in melanoma, cervical cancer, and lung cancer in mouse models of these diseases [45,53]. Also, the reintroduction of sodium butyrate, a metabolite of SCFAs normally produced by Gram-positive bacteria, nullified the improvement in radiotherapy seen with antibiotic treatment [45]. The complete removal of all the gut bacteria has been shown to reduce the efficacy of radiation treatment in breast cancer and melanoma models, while the complete eradication of all the fungi improved this efficacy, possibly due to the opposing effects on immune recruitment [54].

Chemotherapy

Published evidence suggests that the gut microbiome is associated with the outcomes of chemotherapy treatment and its side effects [55]. The purpose of chemotherapy is to disrupt cell division, which triggers apoptosis and, occasionally, an immune response. Some normal cells, which are also subject to cell division, can also be affected, leading to the commonly observed side effects of these therapies. *Fusobacterium nucleatum*, for example, can activate autophagy, promote chemoresistance, and inhibit the caspase pathway [56]. Reducing the gut microbiota by antibiotics has led to reduced effects of chemotherapy drugs such as cisplatin and oxaliplatin in colon cancers and lymphomas [57]. Fecal microbiota transfer (FMT) to mice revealed that *Paraprevotella clara* is associated with responders to the action of oxaliplatin, while *B. fragilis* was linked with non-responders [58]. When focusing on individual metabolites, it was found that indole-3-acetic acid can increase the therapeutic efficacy of FOLFIRINOX (a mix of chemicals containing folinic acid, fluorouracil, irinotecan, and oxaliplatin) in patients with pancreatic ductal adenocarcinoma [59]. Bacteria within the tumor can also influence chemotherapeutic efficacy, as illustrated by the human commensal *E. coli*. These bacteria were found to increase the toxicity of tegafur and mercaptopurine while lowering that of doxorubicin and

mitoxantrone through their influence on the metabolism of these chemotherapeutic agents [60].

Immunotherapy

The gut microbiome can influence the patient's immune response and manipulate the effectiveness of anticancer medicines [61,62]. The three main immune checkpoint therapies targeted by drugs are CTLA-4, PD-1, and PD-L1 [63]. Drugs targeting CTLA-4 were found to be most effective in the presence of the gut microbiome. Recolonization with *B. fragilis*, *Bacteroides thetaiotaomicron*, and *Burkholderia cepacia* (or their recombination) following antibiotic treatment has led to improved anti-CTLA-4 efficacy [64]. Ablation of the gut microbiome with antibiotics has been shown to reduce tumor growth in mouse models of pancreatic ductal adenocarcinoma, particularly when combined with anti-PD-1 immunotherapies [65]. As PD-L1 represents the ligand expressed by tumour cells for the receptor PD-1, which is expressed by T cells, both proteins have been shown to be similarly dependent on the gut microbiome. The use of CAR-T cells is another arm of immunotherapy, and studies have shown that patients receiving broad-spectrum antibiotics were more likely to have worse survival outcomes [66].

The Microbiome Driving New Cancer Treatments

Microbiota-driven cancer therapies have the potential to improve cancer outcomes. These types of treatments are considered briefly in the following sections.

Antibiotics

Antibiotics can alter the microbiota and their metabolism, thus modulating the immune response [17]. The quinolone group of antibiotics eliminates the pathogenic bacteria *Klebsiella pneumoniae* and improves the survival rate of pancreatic cancer patients [67]. Also, mice nebulized with vancomycin/neomycin caused a reduction in the bacterial load of the lungs associated with a reduction in Treg cells and an activation of T and NK cells, leading to a significant decrease in lung metastasis [68]. More recently, CRISPR-Cas-based antimicrobials emerged to eliminate detrimental bacteria while preserving the beneficial microbiome. However, system delivery and safety in avoiding unintended genetic changes remain significant challenges. Additionally, studies have shown that antibiotics may also promote carcinogenesis by promoting chronic inflammation, leading to genotoxicity and weakened immune response, thereby adversely impacting cancer treatment and acting as a double-edged sword [69].

Probiotics

Probiotics are live microorganisms that have been explored for their potential to manipulate the gut microbiome and improve cancer treatment outcomes. Probiotics can enhance the efficacy of cancer therapies, reduce side effects, and potentially prevent cancer development by modulating the gut's microbial

environment [70]. This treatment mainly consists of *Lactobacillus* and *Bifidobacterium* species [17]. Probiotics influence carcinogenesis by regulating gut bacteria and their metabolism and inhibiting the production of carcinogenic substances [71]. They can also inhibit immune response and apoptosis, which ultimately affects cancer progression.

Diet

The food we consume plays a crucial role in shaping our microbiome, generating metabolites that can modulate carcinogenesis and cancer treatment response [72]. The non-digestible part of the diet, prebiotics, can affect the content and function of the microbiome, resulting in beneficial effects in enhancing the effectiveness of cancer therapies, particularly immunotherapies [73]. Examples of prebiotics include the different types of oligosaccharides, inulin, and resistant starch, which are preferentially metabolized by different bacteria into SCFAs, mainly acetate, propionate, and butyrate. Prebiotics can improve resistance against pathogenic bacteria, maintain the integrity of the mucous and colonic epithelial cells, and improve anticancer effects [74,75]. A diet high in processed foods and low in fiber may negatively influence the microbiome and reduce treatment efficacy [76].

Bacteriophage and oncolytic virus therapy

While antibiotics can control bacterial growth, they possess a dysbiotic action on the microbiome and often fail to penetrate biofilms [77]. Bacteriophages (viruses that infect bacteria) and other oncolytic viruses can be used for the treatment of cancer, the former for their ability to attack and eliminate unwanted bacteria, and the latter for killing cancer cells. Bacteriophages can target intratumoral bacteria and cause lysis of these microorganisms to help reduce tumor growth. Bacteriophages can target and eliminate *F. nucleatum*, a bacterium implicated in colorectal cancer, and enhance the activity and effectiveness of chemotherapy [78]. Other viruses might be employed for their ability to target and eliminate cancer cells, as in the case of Talimogene laherparepvec (T-VEC). This product is a genetically engineered herpes simplex virus type 1 that replicates in tumor cells to induce an antitumor response in melanoma [79,80].

Microbial transplantation

Fecal microbial transplantation (FMT) and selective microbial transplantation (SMT) are becoming potential therapies in cancer treatment, particularly in conjunction with immunotherapy [81]. It involves transferring gut bacteria from a healthy donor to a patient to restore/alter the microbial balance of the host and potentially improve the body's immune response to cancer. Microbial transplantation, through regulating the host's immune response, may improve the performance of cancer therapies such as immune checkpoint inhibitors [82]. Several clinical trials are currently taking place investigating the effect of FMT on a variety of cancer types.

Bacterial-based cancer therapy

A few bacteria can naturally target and accumulate in tumours and can be used to deliver therapeutic agents to the cancer cells or stimulate the immune response to eliminate them. Some bacteria can induce immune responses, through the activation of T cells and cytokines, which recognize and eliminate cancer cells [83,84]. *E. coli* was shown to activate T lymphocytes to attack tumour cells, and *Salmonella typhimurium* was found to activate cytokines such as IL-1 β , TNF- α IL-18 [84]. Other bacteria release substances, proteins, or toxins, such as colicin from *E. coli* and microcin E492 from *Klebsiella pneumoniae*, to infiltrate and destroy the cancer cell [85,86]. *Streptococci* bacteria have been proven to clear tumors by releasing polysaccharides, forming a biofilm that represses the attachment of cancer cells to endothelial cells [84]. Additionally, bacteria can serve as carriers to deliver cancer medicine to tumors. Bacteria of the genus *Salmonella* have been shown to express a protein called endostatin, which can block angiogenesis [87].

Conclusions

A large and growing body of research has shown the profound effect of the human microbiome on cancer development, progression, and treatment response. Accumulating evidence revealed that the gut microbiome, in addition to intratumoral bacteria, modulates the immune function and metabolic processes. Dysbiosis, a state of imbalance of the body's microbial community, has been associated with various cancers, pointing to its diagnostic and therapeutic applications. Challenges remain, including the need for better and standardized methodologies for the assessment of the commensal microbial community and any changes that may occur following cancer and its treatment. The microbiome's main value likely lies in its role as a modifying factor influencing risk, progression, and response to cancer therapies. Considering that, future research could note interindividual variations in the microbiome and design personalized microbiome interventions, paving the way for innovative "standalone" therapy or therapy in conjunction with other more established approaches.

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REFERENCES

1. Sepich-Poore GD, Zitvogel L, Straussman R, Hasty J, Wargo JA, Knight R. The microbiome and human cancer. *Science*. 2024;385(6716):eadt2260. doi: 10.1126/science.adt2260.
2. Afzaal M, Saeed F, Shah YA, Hussain M, Rabail R, Socol CT, et al. Human gut microbiota in health and disease: Unveiling the relationship. *Front Microbiol*. 2022;13:999001. doi: 10.3389/fmicb.2022.999001.
3. Sender R, Fuchs S, Milo R. Revised estimates for the number of human and bacteria cells in the body. *PLoS Biol*. 2016;14(8):e1002533. doi: 10.1371/journal.pbio.1002533.
4. Iebba V, Totino V, Gagliardi A, Santangelo F, Cacciotti F, Trancassini M, et al. Eubiosis and dysbiosis: the two sides of the microbiota. *New Microbiol*. 2016;39(1):1-12. PMID: 26922981.
5. Nejman D, Livvyatan I, Fuks G, Gavert N, Zwang Y, Geller LT, et al. The human tumor microbiome is composed of tumor type-specific intracellular bacteria. *Science*. 2020;368(6494):973-980. doi: 10.1126/science.aay9189.
6. Dion MB, Oechslin F, Moineau S. Phage diversity, genomics and phylogeny. *Nat Rev Microbiol*. 2020;18(3):125-138. doi: 10.1038/s41579-019-0311-5.
7. Sekirov I, Russell SL, Antunes LC, Finlay BB. Gut microbiota in health and disease. *Physiol Rev*. 2010;90(3):859-904. doi: 10.1152/physrev.00045.2009.
8. Rajagopala SV, Vashee S, Oldfield LM, Suzuki Y, Venter JC, Telenti A, et al. The human microbiome and cancer. *Cancer Prev Res (Phila)*. 2017;10(4):226-234. doi: 10.1158/1940-6207.CAPR-16-0249.
9. Cullin N, Azevedo Antunes C, Straussman R, Stein-Thoeringer CK, Elinav E. Microbiome and cancer. *Cancer Cell*. 2021;39(10):1317-1341. doi: 10.1016/j.ccell.2021.08.006.
10. IARC monographs on the evaluation of carcinogenic risks to humans 100B. Available at: <https://publications.iarc.fr/Book-And-Report-Series/Iarc-Monographs-On-The-Identification-Of-Carcinogenic-Hazards-To-Humans/Biological-Agents-2012>.
11. Gregory AC, Zablocki O, Zayed AA, Howell A, Bolduc B, Sullivan MB. The gut virome database reveals age-dependent patterns of virome diversity in the human gut. *Cell Host Microbe*. 2020;28(5):724-740.e8. doi: 10.1016/j.chom.2020.08.003.
12. Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nat Med*. 2013;19(11):1423-37. doi: 10.1038/nm.3394.
13. Hou K, Wu ZX, Chen XY, Wang JQ, Zhang D, Xiao C, et al. Microbiota in health and diseases. *Signal Transduct Target Ther*. 2022;7(1):135. doi: 10.1038/s41392-022-00974-4.
14. Zhou P, Hu Y, Wang X, Shen L, Liao X, Zhu Y, et al. Microbiome in cancer: An exploration of carcinogenesis, immune responses and immunotherapy. *Front Immunol*. 2022;13:877939. doi: 10.3389/fimmu.2022.877939.
15. Ding T, Liu C, Li Z. The mycobiome in human cancer: analytical challenges, molecular mechanisms, and therapeutic implications. *Mol Cancer*. 2025;24(1):18. doi: 10.1186/s12943-025-02227-8.
16. Feng M, Cheng X. Parasite-associated cancers (Blood flukes/Liver flukes). *Adv Exp Med Biol*. 2017;1018:193-205. doi: 10.1007/978-981-10-5765-6_12.
17. Chen Z, Guan D, Wang Z, Li X, Dong S, Huang J, et al. Microbiota in cancer: molecular mechanisms and therapeutic interventions. *MedComm (2020)*. 2023;4(6):e417. doi: 10.1002/mco2.417.
18. Barrett M, Hand CK, Shanahan F, Murphy T, O'Toole PW. Mutagenesis by microbe: The role of the microbiota in shaping the cancer genome. *Trends Cancer*. 2020;6(4):277-287. doi: 10.1016/j.trecan.2020.01.019.
19. Wroblewski LE, Peek RM. Helicobacter pylori in gastric carcinogenesis: mechanisms. *Gastroenterol Clin North Am*. 2013;42(2):285-298. doi: 10.1016/j.gtc.2013.01.006.
20. Dzutsev A, Badger JH, Perez-Chanona E, Roy S, Salcedo R, Smith CK, et al. Microbes and cancer. *Annu Rev Immunol*. 2017;35:199-228. doi: 10.1146/annurev-immunol-051116-052133.
21. Machado AM, Figueiredo C, Touati E, Máximo V, Sousa S, Michel V, et al. Helicobacter pylori infection induces genetic instability of nuclear and mitochondrial DNA in gastric cells. *Clin Cancer Res*. 2009;15(9):2995-3002. doi: 10.1158/1078-0432.CCR-08-2686.

22. Mousa WK. The microbiome-product colibactin hits unique cellular targets mediating host-microbe interaction. *Front Pharmacol.* 2022;13:958012. doi: 10.3389/fphar.2022.958012.
23. Kim JJ, Tao H, Carloni E, Leung WK, Graham DY, Sepulveda AR. Helicobacter pylori impairs DNA mismatch repair in gastric epithelial cells. *Gastroenterology.* 2002;123(2):542-553. doi: 10.1053/gast.2002.34751.
24. Hsueh CY, Lau HC, Huang Q, Gong H, Sun J, Cao P, et al. Fusobacterium nucleatum impairs DNA mismatch repair and stability in patients with squamous cell carcinoma of the head and neck. *Cancer.* 2022;128(17):3170-3184. doi: 10.1002/cncr.34338.
25. Lu R, Wu S, Zhang YG, Xia Y, Liu X, Zheng Y, et al. Enteric bacterial protein AvrA promotes colonic tumorigenesis and activates colonic beta-catenin signaling pathway. *Oncogenesis.* 2014;3(6):e105. doi: 10.1038/oncsis.2014.20.
26. Del Bel Belluz L, Guidi R, Pateras IS, Levi L, Mihaljevic B, Rouf SF, et al. The typhoid toxin promotes host survival and the establishment of a persistent asymptomatic infection. *PLoS Pathog.* 2016;12(4):e1005528. doi: 10.1371/journal.ppat.1005528.
27. Scanu T, Spaepen RM, Bakker JM, Pratap CB, Wu LE, Hofland I, et al. Salmonella manipulation of host signaling pathways provokes cellular transformation associated with gallbladder carcinoma. *Cell Host Microbe.* 2015;17(6):763-774. doi: 10.1016/j.chom.2015.05.002.
28. Mu W, Jia Y, Chen X, Li H, Wang Z, Cheng B. Intracellular Porphyromonas gingivalis promotes the proliferation of colorectal cancer cells via the MAPK/ERK signaling pathway. *Front Cell Infect Microbiol.* 2020;10:584798. doi: 10.3389/fcimb.2020.584798.
29. Tannahill GM, Curtis AM, Adamik J, Palsson-McDermott EM, McGettrick AF, Goel G, et al. Succinate is an inflammatory signal that induces IL-1 β through HIF-1 α . *Nature.* 2013;496(7444):238-2342. doi: 10.1038/nature11986.
30. Benítez-Páez A, Gómez Del Pulgar EM, Sanz Y. The glycolytic versatility of Bacteroides uniformis CECT 7771 and its genome response to oligo and polysaccharides. *Front Cell Infect Microbiol.* 2017;7:383. doi: 10.3389/fcimb.2017.00383.
31. Donohoe DR, Holley D, Collins LB, Montgomery SA, Whitmore AC, Hillhouse A, et al. A gnotobiotic mouse model demonstrates that dietary fiber protects against colorectal tumorigenesis in a microbiota- and butyrate-dependent manner. *Cancer Discov.* 2014;4(12):1387-1397. doi: 10.1158/2159-8290.CD-14-0501.
32. Lefebvre P, Cariou B, Lien F, Kuipers F, Staels B. Role of bile acids and bile acid receptors in metabolic regulation. *Physiol Rev.* 2009;89(1):147-191. doi: 10.1152/physrev.00010.2008.
33. Chen Z, Wang Z, Du Y, Shi H, Zhou W. The microbiota and aging microenvironment in pancreatic cancer: Cell origin and fate. *Biochim Biophys Acta Rev Cancer.* 2022;1877(6):188826. doi: 10.1016/j.bbcan.2022.188826.
34. Zheng D, Liwinski T, Elinav E. Interaction between microbiota and immunity in health and disease. *Cell Res.* 2020;30(6):492-506. doi: 10.1038/s41422-020-0332-7.
35. Mima K, Nishihara R, Qian ZR, Cao Y, Sukawa Y, Nowak JA, et al. Fusobacterium nucleatum in colorectal carcinoma tissue and patient prognosis. *Gut.* 2016;65(12):1973-1980. doi: 10.1136/gutjnl-2015-310101.
36. Rubinstein MR, Wang X, Liu W, Hao Y, Cai G, Han YW. Fusobacterium nucleatum promotes colorectal carcinogenesis by modulating E-cadherin/ β -catenin signaling via its FadA adhesin. *Cell Host Microbe.* 2013;14(2):195-206. doi: 10.1016/j.chom.2013.07.012.
37. Kim DJ, Park JH, Franchi L, Backert S, Núñez G. The Cag pathogenicity island and interaction between TLR2/NOD2 and NLRP3 regulate IL-1 β production in Helicobacter pylori infected dendritic cells. *Eur J Immunol.* 2013;43(10):2650-2658. doi: 10.1002/eji.201243281.
38. Grivnennikov SI, Wang K, Mucida D, Stewart CA, Schnabl B, Jauch D, et al. Adenoma-linked barrier defects and microbial products drive IL-23/IL-17-mediated tumour growth. *Nature.* 2012;491(7423):254-258. doi: 10.1038/nature11465.
39. Tan J, McKenzie C, Potamitis M, Thorburn AN, Mackay CR, Macia L. The role of short-chain fatty acids in health and disease. *Adv Immunol.* 2014;121:91-119. doi: 10.1016/B978-0-12-800100-4.00003-9.
40. Xu M, Yamada M, Li M, Liu H, Chen SG, Han YW. FadA from Fusobacterium nucleatum utilizes both secreted and nonsecreted forms for functional oligomerization for attachment and invasion of host cells. *J Biol Chem.* 2007;282(34):25000-25009. doi: 10.1074/jbc.M611567200.
41. Haghi F, Goli E, Mirzaei B, Zeighami H. The association between fecal enterotoxigenic B. fragilis with colorectal cancer. *BMC Cancer.* 2019;19(1):879. doi: 10.1186/s12885-019-6115-1.
42. Ting NL, Lau HC, Yu J. Cancer pharmacomicrobiomics: targeting microbiota to optimise cancer therapy outcomes. *Gut.* 2022;71(7):1412-1425. doi: 10.1136/gutjnl-2021-326264.
43. Bronckaers A, Balzarini J, Liekens S. The cytostatic activity of pyrimidine nucleosides is strongly modulated by Mycoplasma hyorhinis infection: Implications for cancer therapy. *Biochem Pharmacol.* 2008;76(2):188-197. doi: 10.1016/j.bcp.2008.04.019.
44. Mahdy MS, Azmy AF, Dishisha T, Mohamed WR, Ahmed KA, Hassan A, et al. Irinotecan-gut microbiota interactions and the capability of probiotics to mitigate Irinotecan-associated toxicity. *BMC Microbiol.* 2023;23(1):53. doi: 10.1186/s12866-023-02791-3.
45. Uribe-Herranz M, Rafail S, Beghi S, Gil-de-Gómez L, Verginadis I, Bittinger K, et al. Gut microbiota modulate dendritic cell antigen presentation and radiotherapy-induced antitumor immune response. *J Clin Invest.* 2020;130(1):466-479. doi: 10.1172/JCI124332.
46. Kim YS, Kim J, Park SJ. High-throughput 16S rRNA gene sequencing reveals alterations of mouse intestinal microbiota after radiotherapy. *Anaerobe.* 2015;33:1-7. doi: 10.1016/j.anaerobe.2015.01.004.
47. Mager DL, Haffajee AD, Devlin PM, Norris CM, Posner MR, Goodson JM. The salivary microbiota as a diagnostic indicator of oral cancer: a descriptive, non-randomized study of cancer-free and oral squamous cell carcinoma subjects. *J Transl Med.* 2005;3:27. doi: 10.1186/1479-5876-3-27.
48. Farrell JJ, Zhang L, Zhou H, Chia D, Elashoff D, Akin D, et al. Variations of oral microbiota are associated with pancreatic diseases including pancreatic cancer. *Gut.* 2012;61(4):582-588. doi: 10.1136/gutjnl-2011-300784.
49. Yan X, Yang M, Liu J, Gao R, Hu J, Li J, et al. Discovery and validation of potential bacterial biomarkers for lung cancer. *Am J Cancer Res.* 2015;5(10):3111-3122.
50. Liang JQ, Li T, Nakatsu G, Chen YX, Yau TO, Chu E, et al. A novel faecal Lachnospirillum marker for the non-invasive diagnosis of colorectal adenoma and cancer. *Gut.* 2020;69(7):1248-1257. doi: 10.1136/gutjnl-2019-318532.
51. Huang YF, Chen YJ, Fan TC, Chang NC, Chen YJ, Midha MK, et al. Analysis of microbial sequences in plasma cell-free DNA for early-onset breast cancer patients and healthy females. *BMC Med Genomics.* 2018;11(Suppl 1):16. doi: 10.1186/s12920-018-0329-y.
52. Chen H, Ma Y, Xu J, Wang W, Lu H, Quan C, et al. Circulating microbiome DNA as biomarkers for early diagnosis and recurrence of lung cancer. *Cell Rep Med.* 2024;5(4):101499. doi: 10.1016/j.xcrm.2024.101499.
53. Kandalai S, Li H, Zhang N, Peng H, Zheng Q. The human microbiome and cancer: a diagnostic and therapeutic perspective. *Cancer Biol Ther.* 2023;24(1):2240084. doi: 10.1080/15384047.2023.2240084.
54. Shiao SL, Kershaw KM, Limon JJ, You S, Yoon J, Ko EY, et al. Commensal bacteria and fungi differentially regulate tumor responses to radiation therapy. *Cancer Cell.* 2021;39(9):1202-1213. doi: 10.1016/j.ccell.2021.07.002.
55. Oh B, Boyle F, Pavlakis N, Clarke S, Guminski A, Eade T, et al. Emerging evidence of the gut microbiome in chemotherapy: A clinical review. *Front Oncol.* 2021;11:706331. doi: 10.3389/fonc.2021.706331.
56. Li S, Zhu S, Yu J. The role of gut microbiota and metabolites in cancer chemotherapy. *J Adv Res.* 2024;64:223-235. doi: 10.1016/j.jare.2023.11.027.
57. Iida N, Dzutsev A, Stewart CA, Smith L, Bouladoux N, Weingarten RA, et al. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. *Science.* 2013;342(6161):967-970. doi: 10.1126/science.1240527.
58. Roberti MP, Yonekura S, Duong CPM, Picard M, Ferrere G, Tidjani Alou M, et al. Chemotherapy-induced ileal crypt apoptosis and the ileal microbiome shape immunosurveillance and prognosis of proximal colon cancer. *Nat Med.* 2020;26(6):919-931. doi: 10.1038/s41591-020-0882-8.

59. Tintelnot J, Xu Y, Lesker TR, Schönlein M, Konczalla L, Giannou AD, et al. *Nature*. 2023;615(7950):168-174. doi: 10.1038/s41586-023-05728-y.
60. Lehouritis P, Cummins J, Stanton M, Murphy CT, McCarthy FO, Reid G, et al. Local bacteria affect the efficacy of chemotherapeutic drugs. *Sci Rep*. 2015;5:14554. doi: 10.1038/srep14554.
61. Zhang M, Liu J, Xia Q. Role of gut microbiome in cancer immunotherapy: from predictive biomarker to therapeutic target. *Exp Hematol Oncol*. 2023;12(1):84. doi: 10.1186/s40164-023-00442-x.
62. Elkrief A, Pidgeon R, Maleki Vareki S, Messaoudene M, Castagner B, Routy B. The gut microbiome as a target in cancer immunotherapy: opportunities and challenges for drug development. *Nat Rev Drug Discov*. 2025. doi: 10.1038/s41573-025-01211-7.
63. Kang X, Lau HC, Yu J. Modulating gut microbiome in cancer immunotherapy: Harnessing microbes to enhance treatment efficacy. *Cell Rep Med*. 2024;5(4):101478. doi: 10.1016/j.xcrm.2024.101478.
64. Vétizou M, Pitt JM, Daillère R, Lepage P, Waldschmitt N, Flament C, et al. Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. *Science*. 2015;350(6264):1079-1084. doi: 10.1126/science.aad1329.
65. Pushalkar S, Hundeyin M, Daley D, Zambirinis CP, Kurz E, Mishra A, et al. The pancreatic cancer microbiome promotes oncogenesis by induction of innate and adaptive immune suppression. *Cancer Discov*. 2018;8(4):403-416. doi: 10.1158/2159-8290.CD-17-1134.
66. Stein-Thoeringer CK, Saini NY, Zamir E, Blumenberg V, Schubert ML, Mor U, et al. A non-antibiotic-disrupted gut microbiome is associated with clinical responses to CD19-CAR-T cell cancer immunotherapy. *Nat Med*. 2023;29(4):906-916. doi: 10.1038/s41591-023-02234-6.
67. Weniger M, Hank T, Qadan M, Ciprani D, Michelakos T, Niess H, et al. Influence of *Klebsiella pneumoniae* and quinolone treatment on prognosis in patients with pancreatic cancer. *Br J Surg*. 2021;108(6):709-716. doi: 10.1002/bjs.12003.
68. Le Noci V, Guglielmetti S, Arioli S, Camisaschi C, Bianchi F, Sommariva M, et al. Modulation of pulmonary microbiota by antibiotic or probiotic aerosol therapy: A strategy to promote immunosurveillance against lung metastases. *Cell Rep*. 2018;24(13):3528-3538. doi: 10.1016/j.celrep.2018.08.090.
69. Gao Y, Shang Q, Li W, Guo W, Stojadinovic A, Mannion C, et al. Antibiotics for cancer treatment: A double-edged sword. *J Cancer*. 2020;11(17):5135-5149. doi: 10.7150/jca.47470.
70. Wang Z, Li L, Wang S, Wei J, Qu L, Pan L, et al. The role of the gut microbiota and probiotics associated with microbial metabolisms in cancer prevention and therapy. *Front Pharmacol*. 2022;13:1025860. doi: 10.3389/fphar.2022.1025860.
71. Lili Z, Junyan W, Hongfei Z, Baoqing Z, Bolin Z. Detoxification of cancerogenic compounds by lactic acid bacteria strains. *Crit Rev Food Sci Nutr*. 2018;58(16):2727-2742. doi: 10.1080/10408398.2017.1339665.
72. Rizvi ZA, Dalal R, Sadhu S, Kumar Y, Kumar S, Gupta SK, et al. High-salt diet mediates interplay between NK cells and gut microbiota to induce potent tumor immunity. *Sci Adv*. 2021;7(37):eabg5016. doi: 10.1126/sciadv.abg5016.
73. Bindels LB, Delzenne NM, Cani PD, Walter J. Towards a more comprehensive concept for prebiotics. *Nat Rev Gastroenterol Hepatol*. 2015;12(5):303-310. doi: 10.1038/nrgastro.2015.47.
74. Verspreet J, Damen B, Broekaert WF, Verbeke K, Delcour JA, Courtin CM. A critical look at prebiotics within the dietary fiber concept. *Annu Rev Food Sci Technol*. 2016;7:167-190. doi: 10.1146/annurev-food-081315-032749.
75. Li Y, Elmén L, Segota I, Xian Y, Tinoco R, Feng Y, et al. Prebiotic-Induced Anti-tumor Immunity Attenuates Tumor Growth. *Cell Rep*. 2020;30(6):1753-1766. doi: 10.1016/j.celrep.2020.01.035.
76. Nguyen NA, Jiang Y, McQuade JL. Eating away cancer: the potential of diet and the microbiome for shaping immunotherapy outcome. *Front Immunol*. 2024;15:1409414. doi: 10.3389/fimmu.2024.1409414.
77. Kabwe M, Dashper S, Bachrach G, Tucci J. Bacteriophage manipulation of the microbiome associated with tumour microenvironments-can this improve cancer therapeutic response? *FEMS Microbiol Rev*. 2021;45(5):fuab017. doi: 10.1093/femsre/fuab017.
78. Guo C, An Q, Zhang LY, Wei XD, Xu J, Yu JY, et al. Intratumoral microbiota as cancer therapeutic target. *Aging Med (Milton)*. 2024;7(5):636-644. doi: 10.1002/agm2.12359.
79. Greig SL. Talimogene Laherparepvec: First global approval. *Drugs*. 2016;76(1):147-154. doi: 10.1007/s40265-015-0522-7.
80. Al-Janabi II. Cancer Vaccines: Another aspect to immunotherapy. *Al-Rafidain J Med Sci*. 2024;7(1):34-45. doi: 10.54133/ajms.v7i1.1057.
81. Wu X, Zhang T, Chen X, Ji G, Zhang F. Microbiota transplantation: Targeting cancer treatment. *Cancer Lett*. 2019;452:144-151. doi: 10.1016/j.canlet.2019.03.010.
82. Yang Y, An Y, Dong Y, Chu Q, Wei J, Wang B, et al. Fecal microbiota transplantation: no longer cinderella in tumour immunotherapy. *EBioMedicine*. 2024;100:104967. doi: 10.1016/j.ebiom.2024.104967.
83. Kucerova P, Cervinkova M. Spontaneous regression of tumour and the role of microbial infection--possibilities for cancer treatment. *Anticancer Drugs*. 2016;27(4):269-277. doi: 10.1097/CAD.0000000000000337.
84. Mills H, Acquah R, Tang N, Cheung L, Klenk S, Glassen R, et al. The Use of Bacteria in Cancer Treatment: A Review from the Perspective of Cellular Microbiology. *Emerg Med Int*. 2022 Aug 8;2022:8127137. doi: 10.1155/2022/8127137.
85. Chumchalová J, Smarda J. Human tumor cells are selectively inhibited by colicins. *Folia Microbiol (Praha)*. 2003;48(1):111-115. doi: 10.1007/BF02931286.
86. Villarante KI, Elegado FB, Iwatani S, Zendo T, Sonomoto K, de Guzman EE. Purification, characterization and in vitro cytotoxicity of the bacteriocin from *Pediococcus acidilactici* K2a2-3 against human colon adenocarcinoma (HT29) and human cervical carcinoma (HeLa) cells. *World J Microbiol Biotechnol*. 2011;27:975-980. doi: 10.1007/s11274-010-0541-1.
87. Aganja RP, Sivasankar C, Senevirathne A, Lee JH. Salmonella as a promising curative tool against cancer. *Pharmaceutics*. 2022;14(10):2100. doi: 10.3390/pharmaceutics14102100.